

## Synthesis and Cytotoxicity of Sempervirine and Analogues

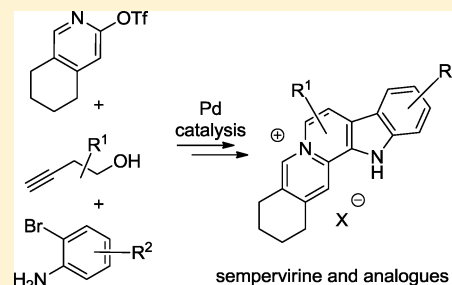
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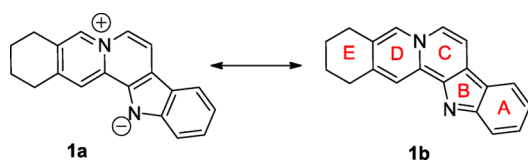
### Supporting Information

**ABSTRACT:** Sempervirine and analogues were synthesized using a route featuring Sonogashira and Larock Pd-catalyzed reactions. Structure–activity relationships were investigated using three human cancer cell lines. 10-Fluorosempervirine is the most potently cytotoxic member of the family yet described.



Many natural products and their derivatives are important therapeutic agents, particularly in the treatment of infections and cancers.<sup>1</sup> Modern synthetic methods, especially those not relying upon the intrinsic reactivity of groups present in the natural material, have lowered the barrier to conducting SAR studies in natural product scaffolds to improve efficacy and therapeutic index. We recently reported a general approach to the synthesis of tetracyclic and pentacyclic alkaloids<sup>2,3</sup> featuring highly substrate-tolerant Sonogashira<sup>3</sup> and Larock<sup>4</sup> reactions. Here we report using the route for an SAR study in the sempervirine series.

Sempervirine has attracted significant interest since the pioneering structure elucidation<sup>5</sup> and synthetic work<sup>6</sup> of Woodward, who proposed its canonical forms (Figure 1).



**Figure 1.** Sempervirine canonical forms and ring numbering (Woodward and Witkop, 1949).<sup>5</sup>

Several elegant syntheses have been reported, notably by Swan,<sup>7</sup> Westphal,<sup>8</sup> Potts,<sup>9</sup> and Gribble.<sup>10</sup> It has been targeted due to its antitumor activity, arising from topoisomerase 1 inhibition, DNA intercalation, and other mechanisms.<sup>11</sup> Routes to sempervirine generally involve moderately long synthetic sequences, proceed with a modest overall yield, and have limitations in scope for preparing the natural material or a small set of analogues.

Previous sempervirine analogues prepared include Woodward's indole *N*-methyl derivative<sup>6</sup> and Lipińska's three analogues, wherein the E-ring (Figure 1) was replaced by fused cyclopentyl, cycloheptyl, and cyclooctyl rings, with each product obtained in six steps in about 4% overall yield, using a

route featuring an inverse electron demand Diels–Alder reaction between a 1,2,4-triazine diene and a cyclic enamine.<sup>12</sup> Lipińska later reported four C10 methoxy analogues having five-, six-, seven-, and eight-membered E-rings, in 2–10% yield by a route using a Fischer indole synthesis<sup>12</sup> and Gribble's method<sup>10</sup> for C-ring construction. Malhotra et al. used a microwave-assisted version of Westphal's method<sup>8</sup> to prepare the C11 methoxy analogue in 32% yield,<sup>13</sup> a compound previously made by Huebner et al.<sup>14</sup> Malhotra et al. also prepared a compound in 42% yield, wherein the E-ring was replaced with two phenyl groups.<sup>13</sup> To our knowledge, the biological activity for these compounds is unreported.

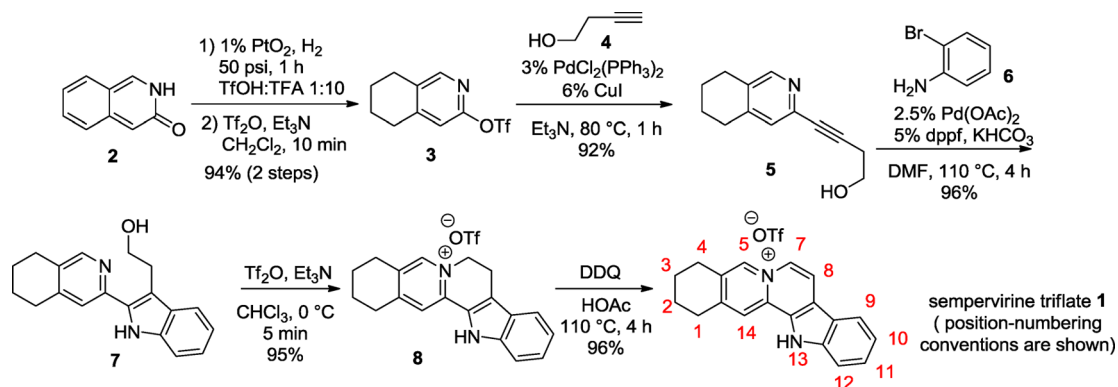
Our route uses an initial regioselective semireduction of 3-isosquinolone (**2**, Scheme 1). Literature reduction methods required excesses of expensive and harsh reagents (SbF<sub>3</sub> or SbF<sub>5</sub> in CF<sub>3</sub>SO<sub>3</sub>H).<sup>15</sup> In a solvent survey using 1 mol % of PtO<sub>2</sub>, acetic acid gave unwanted pyridone ring saturation. Adding TFA favored the desired phenyl ring reduction, with 10:1 TFA/triflic acid giving full regioselectivity. The triflate **3** was then prepared (94% yield, two steps) and used in a Sonogashira reaction (92%) followed by a Larock indole synthesis reaction that proceeded with high regioselectivity<sup>16</sup> and then a triflate-promoted cyclization (91%, two steps).<sup>2</sup> DDQ oxidation provided sempervirine triflate **1** (96%). The route is six steps and 76% yield, with intermediate **8** and the natural product purified by precipitation. Given the efficiency, ease of purification, and expected substrate tolerances, we felt that an SAR study for sempervirine was feasible.

Altering ring size within the core is a drastic change not generally possible by natural product modification. A seven-membered ring analogue of dihydrosempervirine was prepared in three steps and 60% yield from triflate **3** (Scheme 2). In the Larock cyclization, catalyst loading was increased 2-fold relative

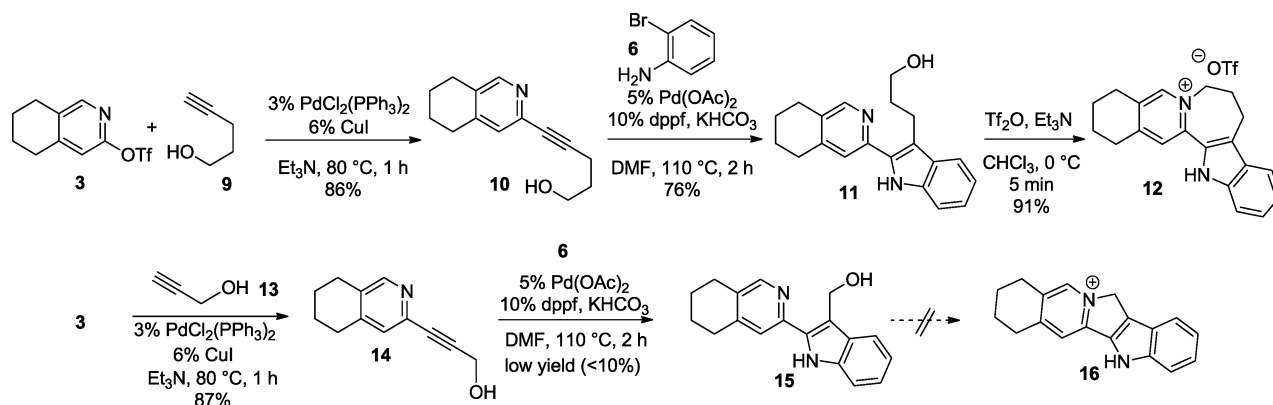
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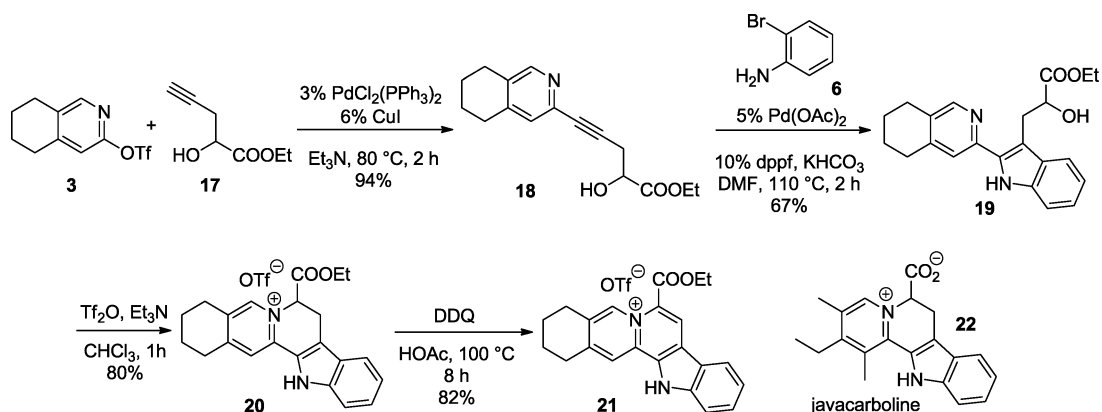
Scheme 1. Synthesis of Sempervirine Triflate



Scheme 2. Approaches to Seven- and Five-Membered Ring Analogues



Scheme 3. C-Ring Carboethoxy-Substituted Analogues



to the Scheme 1 protocol. The lower yield seen for a longer alkyl chain (compound 10 vs 5) suggests hydroxyl group chelation in the alkyne palladation step. The Larock reaction to access the five-membered ring analogue 16 (Scheme 2) was unsuccessful, perhaps due to poor coordination by the propargylic alcohol group and/or gramine-like instability of product 15.

Structural diversity in the central ring is possible: a C7 substituent can be installed by using hydroxyester 17<sup>17</sup> to give tetracyclic esters 20 and 21 (Scheme 3). This core ring system and pattern of substitution are similar to that of javacarboline (22).

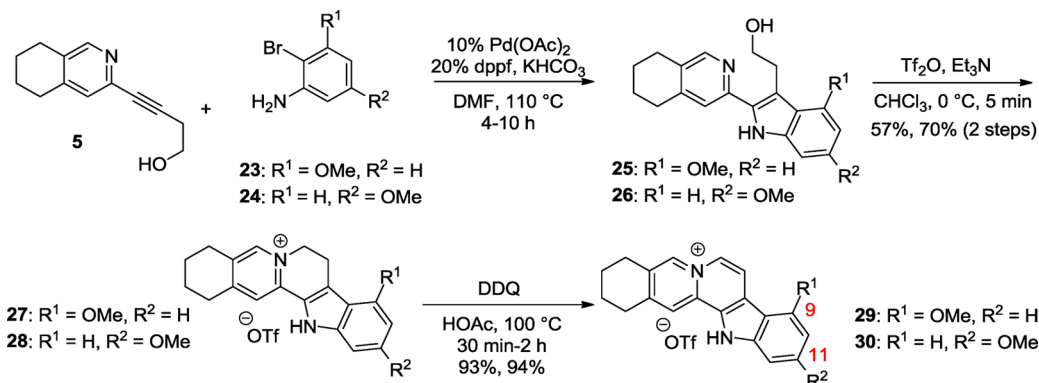
Using electron-rich bromoanilines permits access to electron-rich sempervirine analogues (29 and 30, Scheme 4). Here,

intermediates 25 and 26 were not purified, and salts 27–30 were isolated by precipitation. Compound 30 in another salt form has been described.<sup>13</sup>

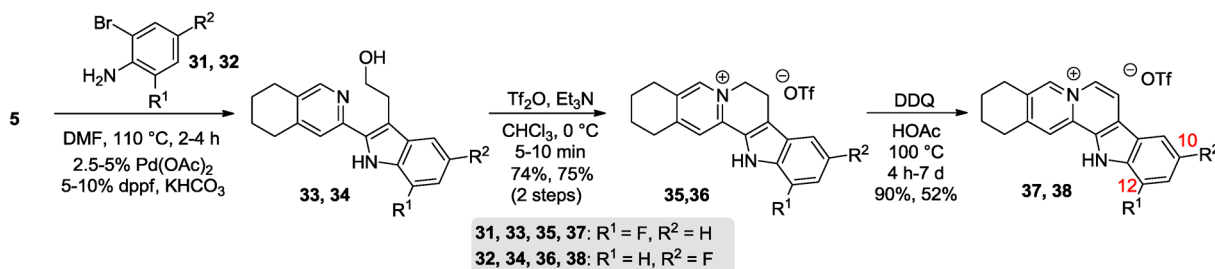
Electron-deficient derivatives are accessible (37 and 38, Scheme 5). For 38, the DDQ aromatization reaction required 7 days and excess oxidant. As another variation, isoquinolinone 2 semireduction can be omitted, providing the aromatic E-ring analogue 43 (Scheme 6).

Sempervirine analogues with substituents in the A-, C-, and/or E-ring are thus accessible. Presumably, the use of C1- or C4-substituted analogues of isoquinoline 2 would also allow access to D-ring analogues (our previous paper described D-ring aza analogues).<sup>2</sup> The B-ring, with only one site for substitution, can likely also be modified by N-alkylation of an intermediate or the

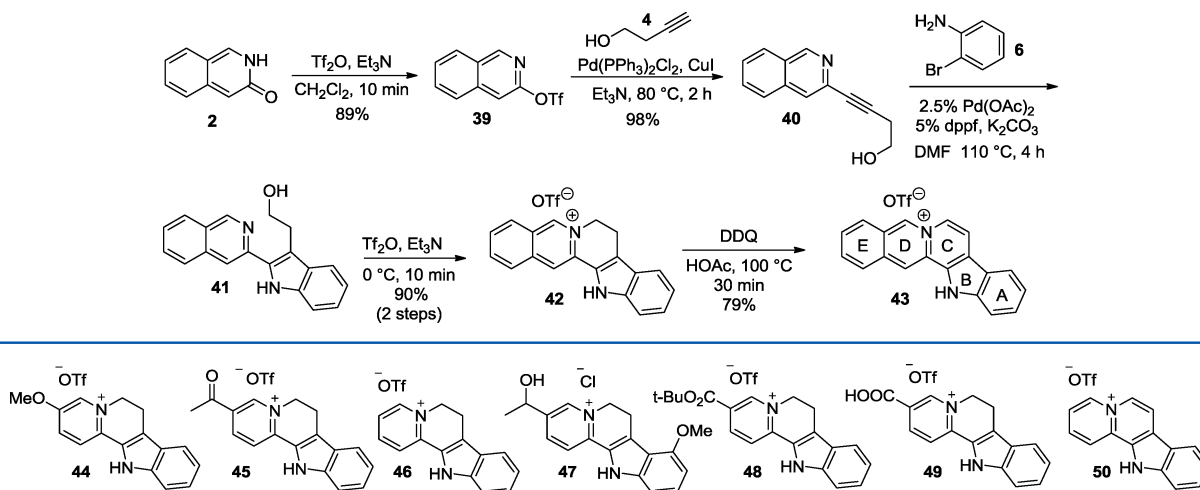
## Scheme 4. A-Ring Methoxy-Substituted Analogues



## Scheme 5. Fluorinated Analogues



## Scheme 6. Aromatic Analogues

Figure 2. Additional analogues tested.<sup>2</sup>

final product. The ability to modify rings A–E of sempervirine by one general method illustrates the versatility and utility of the Larock and Sonogashira reactions.

Because sempervirine has multiple mechanisms of antitumor activity,<sup>11</sup> a single biochemical assay is inadequate to assess potency. We conducted standard whole-cell antitumor viability MTT assays<sup>18</sup> in three sempervirine-sensitive human tumor lines: Raji Burkitt lymphoma, MDA-MB-231 breast cancer, and HeLa cervical cancer cells. Compounds **44**–**50** (Figure 2)<sup>2</sup> were also evaluated. Growth inhibition EC<sub>50</sub> estimates were determined using a 6-point dilution protocol.<sup>18</sup> Active compounds (estimated EC<sub>50</sub> < 3 μM) were re-evaluated using a 12-point protocol.

Growth inhibition curves for sempervirine and two more potent analogues are shown in Figure 3. A 2.3–3.4-fold enhancement of potency for **38** relative to that for sempervirine **1** was found. Compound **38** is, to our knowledge, the most cytotoxic sempervirine analogue known. Enhanced potency and improved pharmacokinetic properties are likely required for animal use of **38**.<sup>19</sup> Growth inhibition results for all other less active compounds are shown in Table 1, based upon the 6-point assay.<sup>18</sup> Compounds with a nonaromatic central ring were inactive at 10 μM (**8**, **12**, **20**, **27**, **28**, **35**, **36**, **42**, and **44**–**49**), thus extended planarity is essential for activity.

In conclusion, these methods allow extensive modifications to sempervirine, with 10-fluorosempervirine (**38**) being the most potently cytotoxic analogue yet described.

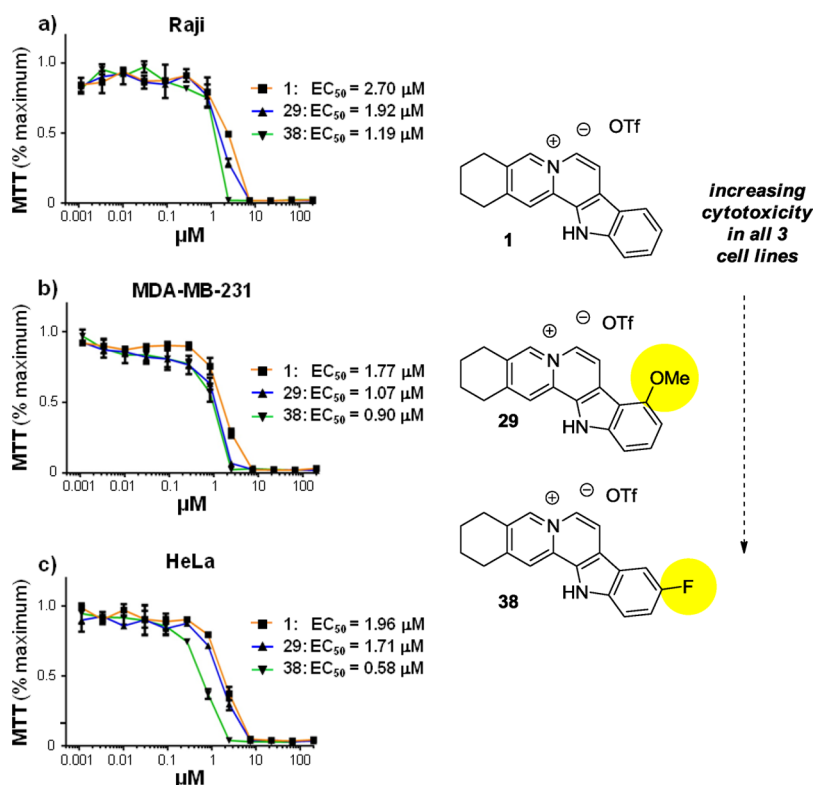


Figure 3. Growth inhibition by the most potent compounds in the series.

Table 1. Estimated Growth Inhibition for All Other Compounds Tested, 6-Point MTT Assay

compound(s)	Raji EC <sub>50</sub> (μM)	MDA-MB-231 EC <sub>50</sub> (μM)	HeLa EC <sub>50</sub> (μM)
8, 12, 20, 27, 28, 35, 36, 42, 44, 45, 46, 47, 48, 49	>10	>10	>10
21, 30, 37, 43	3–10	3–10	3–10
50	3–10	3–10	>10

## EXPERIMENTAL SECTION

**General Methods.** Infrared (IR) spectra were collected on an FT-IR spectrometer. <sup>1</sup>H NMR spectra were recorded at 400 MHz in parts per million (ppm) downfield from an internal standard, CHCl<sub>3</sub> (δ 7.26) or DMSO (δ 2.54). <sup>13</sup>C NMR spectra were recorded at 100 MHz in parts per million (ppm) downfield from an internal standard, CHCl<sub>3</sub> (δ 77.36) or DMSO (δ 40.45). High-resolution mass spectra (HRMS) were recorded on a TOF LC/MS spectrometer. Reagents and anhydrous solvents used were obtained from commercial vendors. Flash column chromatography was performed to purify compounds as indicated, using 60 Å mesh silica columns and automated instruments.

**General Procedure for DDQ Oxidation Reactions. Preparation of Sempervirine Triflate (1).** A solution of **8** (106 mg, 0.25 mmol) and 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (85 mg, 0.375 mmol) in acetic acid (1 mL) was stirred at 110 °C for 4 h. After being cooled to room temperature, the resulting precipitate was filtered and recrystallized with acetic acid, providing **1** (101 mg, 96%) as a yellow solid: IR (neat cm<sup>-1</sup>) 3186, 1651, 1222, 1028, 745; <sup>1</sup>H NMR (400 MHz, DMSO) δ 13.29 (s, 1H), 9.27 (s, 1H), 8.89 (d, J = 6.8 Hz, 1H), 8.72 (d, J = 6.8 Hz, 1H), 8.69 (s, 1H), 8.43 (d, J = 8.0 Hz, 1H), 7.87 (d, J = 8.0 Hz, 1H), 7.75 (t, J = 7.6 Hz, 1H), 7.51 (t, J = 7.6 Hz, 1H), 3.20 (br, 2H), 3.04 (br, 2H), 1.95 (br, 4H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 149.9, 141.2, 135.3, 133.6, 130.8, 130.0, 129.6, 126.6, 122.4, 122.3, 121.6, 121.3, 119.9, 116.6, 113.3, 29.6, 26.6, 22.1, 22.0; HRMS (ES) *m/e* calcd for C<sub>19</sub>H<sub>17</sub>N<sub>2</sub> (M - OTf)<sup>+</sup> 273.1392, found 273.1399.

**5,6,7,8-Tetrahydroisoquinolin-3-yl Trifluoromethanesulfonate (3).** A solution of isoquinolin-3-ol (**2**) (2 g, 13.79 mmol) and PtO<sub>2</sub> (30 mg, 0.132 mmol) in trifluoroacetic acid (10 mL) and triflic acid (1 mL) was stirred under 50 psi H<sub>2</sub> for 1 h. The mixture was filtered, and to the solution was added ice water. The resulting mixture was made basic (pH 8) with saturated aqueous Na<sub>2</sub>CO<sub>3</sub> and extracted with ethyl acetate. The extracts were combined and evaporated to provide 5,6,7,8-tetrahydroisoquinolin-3-ol (2 g, 98%) as a solid. To a solution of 5,6,7,8-tetrahydroisoquinolin-3-ol (200 mg, 1.343 mmol) and Et<sub>3</sub>N (163 mg, 1.612 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) at 0 °C was then added trifluoromethanesulfonic anhydride (0.245 mL, 1.447 mmol) slowly, followed by stirring for 10 min. Water was added, and the CH<sub>2</sub>Cl<sub>2</sub> layer was separated and then filtered through a pad of silica gel. Evaporation of the CH<sub>2</sub>Cl<sub>2</sub> provided compound **3** (360 mg, 96%) as an oil: IR (neat cm<sup>-1</sup>) 2942, 1416, 1201, 927, 834; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.05 (s, 1H), 6.87 (s, 1H), 2.83–2.80 (m, 2H), 2.78–2.75 (m, 2H), 1.87–1.78 (m, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 154.2, 152.3, 148.9, 134.6, 120.5 (CF<sub>3</sub>), 117.4 (CF<sub>3</sub>), 115.0, 29.4, 26.1, 22.5, 22.1; HRMS (ES) *m/e* calcd for C<sub>10</sub>H<sub>11</sub>F<sub>3</sub>NO<sub>3</sub>S (M + H)<sup>+</sup> 282.0406, found 282.0408.

**General Procedure for Sonogashira Coupling Reactions. 4-(5,6,7,8-Tetrahydroisoquinolin-3-yl)but-3-yn-1-ol (5).** A solution of 5,6,7,8-tetrahydroisoquinolin-3-yl triflate (**3**) (1.69 g, 6.0 mmol), 3-butyn-1-ol (**4**) (504 mg, 7.2 mmol), PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> (127 mg, 0.18 mmol), and CuI (69 mg, 0.36 mmol) in Et<sub>3</sub>N (6 mL) was stirred at 80 °C for 1 h. Silica gel was added, and the resulting mixture was evaporated and separated by flash chromatography (SiO<sub>2</sub>, EtOAc), providing compound **5** (1100 mg, 92%) as a solid: IR (neat cm<sup>-1</sup>) 3183, 2937, 1597, 1053, 695; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.15 (s, 1H), 7.04 (s, 1H), 3.85 (t, J = 6.4 Hz, 2H), 2.68–2.65 (m, 6H), 1.77–1.75 (m, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 150.3, 146.9, 140.0, 132.8, 127.1, 87.1, 81.8, 61.0, 28.7, 26.4, 24.1, 22.6, 22.4; HRMS (ES) *m/e* calcd for C<sub>13</sub>H<sub>16</sub>NO (M + H)<sup>+</sup> 202.1226, found 202.1229.

**General Procedure for Larock Indole Synthesis Reactions. 2-(2-(5,6,7,8-Tetrahydroisoquinolin-3-yl)-1H-indol-3-yl)ethanol (7).** 2-Bromoaniline (**6**) (52 mg, 0.3 mmol), **5** (62 mg, 0.3 mmol), Pd(OAc)<sub>2</sub> (1.8 mg, 0.0075 mmol), 1,1'-bis(diphenylphosphino)ferrocene (8.4 mg, 0.015 mmol), and KHCO<sub>3</sub> (90 mg, 0.9 mmol)

were added to dry and degassed DMF (2 mL). The solution was heated for 4 h at 110 °C, after which time the reaction was complete, as determined by LCMS analysis. Water was added, and the mixture was extracted with ethyl acetate. The organic extracts were combined, dried, and purified by flash chromatography (SiO<sub>2</sub>, hexanes/EtOAc = 1:1), providing compound 7 (84 mg, 96%) as a solid: IR (neat cm<sup>-1</sup>) 3182, 2925, 1603, 1042, 737; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.40 (br, 1H), 8.13 (s, 1H), 7.56 (d, *J* = 8.0 Hz, 1H), 7.44 (s, 1H), 7.38 (d, *J* = 8.0 Hz, 1H), 7.21 (t, *J* = 7.2 Hz, 1H), 7.11 (t, *J* = 7.2 Hz, 1H), 4.05 (t, *J* = 6.0 Hz, 2H), 3.26 (t, *J* = 6.0 Hz, 2H), 2.80 (br, 2H), 2.67 (br, 2H), 1.80 (br, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 148.8, 147.7, 147.4, 136.5, 134.1, 131.7, 129.3, 123.0, 121.3, 119.7, 118.9, 112.6, 111.8, 64.0, 29.2, 27.5, 26.3, 22.7, 22.5; HRMS (ES) *m/e* calcd for C<sub>19</sub>H<sub>21</sub>N<sub>2</sub>O (M + H)<sup>+</sup> 293.1648, found 293.1656.

**General Procedure for Triflate Cyclization Reactions.** *7,8-Dihydrosempervirine (8)*. To a solution of compound 7 (44 mg, 0.15 mmol) and Et<sub>3</sub>N (24 mg, 0.225 mmol) in dry CHCl<sub>3</sub> (1.4 mL) at 0 °C was slowly added trifluoromethanesulfonic anhydride (34 μL, 0.18 mmol). After being stirred for an additional 5 min, the resulting yellow precipitate was collected by filtration, and the solid was washed with CHCl<sub>3</sub> and then identified as compound 8 as a yellow solid (60.2 mg, 95%): IR (neat cm<sup>-1</sup>) 3297, 1558, 1149, 1031, 748; <sup>1</sup>H NMR (400 MHz, DMSO) δ 12.21 (s, 1H), 8.81 (s, 1H), 8.01 (s, 1H), 7.75 (d, *J* = 8.0 Hz, 1H), 7.57 (d, *J* = 8.0 Hz, 1H), 7.40 (t, *J* = 7.2 Hz, 1H), 7.22 (t, *J* = 7.2 Hz, 1H), 4.83 (br, 2H), 3.38 (br, 2H), 3.05 (br, 2H), 2.88 (br, 2H), 1.86 (br, 4H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 158.2, 145.4, 140.2, 139.8, 134.4, 126.6, 125.9, 125.7, 121.5, 121.2, 121.0, 116.9, 113.4, 55.9, 29.8, 26.4, 21.9, 21.8, 19.8; HRMS (ES) *m/e* calcd for C<sub>19</sub>H<sub>19</sub>N<sub>2</sub> (M - OTf)<sup>+</sup> 275.1548, found 275.1553.

*5-(5,6,7,8-Tetrahydroisoquinolin-3-yl)pent-4-yn-1-ol (10)*. By the general Sonogashira coupling procedure, 5,6,7,8-tetrahydroisoquinolin-3-yl triflate (3) (141 mg, 0.5 mmol), 4-pentyn-1-ol (9) (51 mg, 0.6 mmol), PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> (11 mg, 0.015 mmol), and CuI (6 mg, 0.03 mmol) in Et<sub>3</sub>N (1 mL) at 80 °C for 1 h provided compound 10 (92 mg, 86%) as an oil after flash chromatography (SiO<sub>2</sub>, EtOAc): IR (neat cm<sup>-1</sup>) 3280, 2931, 1595, 1473, 1060; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.18 (s, 1H), 7.05 (s, 1H), 3.81 (t, *J* = 6.0 Hz, 2H), 2.78 (br, 1H), 2.71–2.66 (m, 4H), 2.55 (t, *J* = 7.2 Hz, 2H), 1.89–1.82 (m, 2H), 1.80–1.73 (m, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 150.6, 146.7, 140.4, 132.6, 127.2, 89.1, 81.0, 61.7, 31.5, 28.7, 26.4, 22.7, 22.5, 16.20; HRMS (ES) *m/e* calcd for C<sub>14</sub>H<sub>18</sub>NO (M + H)<sup>+</sup> 216.1383, found 216.1385.

*3-(2-(5,6,7,8-Tetrahydroisoquinolin-3-yl)-1H-indol-3-yl)propan-1-ol (11)*. By the general Larock indole synthesis procedure, 2-bromoaniline (6) (48 mg, 0.28 mmol), alkyne 10 (60 mg, 0.28 mmol), Pd(OAc)<sub>2</sub> (3.2 mg, 0.014 mmol), 1,1'-bis(diphenylphosphino)ferrocene (15.5 mg, 0.028 mmol), and KHCO<sub>3</sub> (84 mg, 0.84 mmol) in DMF (1.4 mL) for 2 h provided 11 (65 mg, 76%) as an oil after flash chromatography (SiO<sub>2</sub>, hexanes/EtOAc = 1:1): IR (neat cm<sup>-1</sup>) 2931, 1604, 1436, 955, 734; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.18 (s, 1H), 8.20 (s, 1H), 7.65 (d, *J* = 7.6 Hz, 1H), 7.38 (d, *J* = 7.6 Hz, 1H), 7.34 (s, 1H), 7.24 (td, *J* = 6.8 and 0.8 Hz, 1H), 7.14 (td, *J* = 8.0 and 0.8 Hz, 1H), 5.58 (br, 1H), 3.62 (t, *J* = 5.6 Hz, 2H), 3.26 (t, *J* = 6.4 Hz, 2H), 2.72–2.65 (m, 4H), 2.10–2.04 (m, 2H), 1.81–1.73 (m, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 149.5, 148.5, 147.9, 136.6, 133.2, 131.9, 129.6, 123.2, 122.1, 119.8, 119.7, 114.4, 111.5, 60.1, 32.2, 29.2, 26.2, 22.7, 22.6, 20.6; HRMS (ES) *m/e* calcd for C<sub>20</sub>H<sub>23</sub>N<sub>2</sub>O (M + H)<sup>+</sup> 307.1805, found 307.1804.

*1,2,3,4,7,8,9,14-Octahydroindolo[2',3':3,4]azepino[1,2-b]isoquinolin-6-ium Triflate (12)*. By the general triflate cyclization procedure, compound 11 (50 mg, 0.163 mmol), Et<sub>3</sub>N (25 mg, 0.245 mmol), and trifluoromethanesulfonic anhydride (33 μL, 0.196 mmol) in dry CHCl<sub>3</sub> (1 mL) provided 12 (65 mg, 91%) as a yellow solid: IR (neat cm<sup>-1</sup>) 3302, 1556, 1154, 1030, 745; <sup>1</sup>H NMR (400 MHz, DMSO) δ 11.85 (s, 1H), 8.85 (s, 1H), 8.17 (s, 1H), 7.72 (d, *J* = 8.0 Hz, 1H), 7.55 (d, *J* = 8.0 Hz, 1H), 7.42 (t, *J* = 7.6 Hz, 1H), 7.20 (t, *J* = 7.6 Hz, 1H), 4.71–4.69 (m, 2H), 3.24 (t, *J* = 6.8 Hz, 2H), 3.07 (br, 2H), 2.90 (br, 2H), 2.42 (br, 2H), 1.88 (br, 4H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 157.1, 145.6, 143.5, 138.7, 135.3, 128.8, 126.9, 126.7, 124.7, 122.1, 121.1, 121.0, 112.9, 60.3, 29.7, 26.6, 26.3, 24.9, 21.92,

21.89; HRMS (ES) *m/e* calcd for C<sub>20</sub>H<sub>21</sub>N<sub>2</sub> (M - OTf)<sup>+</sup> 289.1705, found 289.1709.

*3-(5,6,7,8-Tetrahydroisoquinolin-3-yl)prop-2-yn-1-ol (14)*. By the general Sonogashira coupling procedure, 5,6,7,8-tetrahydroisoquinolin-3-yl triflate 3 (140 mg, 0.5 mmol), 2-propyn-1-ol 13 (34 mg, 0.6 mmol), PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> (11 mg, 0.015 mmol), and CuI (6 mg, 0.03 mmol) in Et<sub>3</sub>N (1 mL) at 80 °C for 1 h provided 14 (81 mg, 87%) as a solid after flash chromatography (SiO<sub>2</sub>, EtOAc): IR (neat cm<sup>-1</sup>) 3143, 2934, 1041, 974, 868; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.21 (s, 1H), 7.13 (s, 1H), 4.51 (s, 2H), 3.84 (br, 1H), 2.73–2.68 (m, 4H), 1.83–1.75 (m, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 150.6, 147.1, 139.5, 133.4, 127.6, 87.6, 84.6, 51.4, 28.8, 26.5, 22.7, 22.4; HRMS (ES) *m/e* calcd for C<sub>12</sub>H<sub>14</sub>NO (M + H)<sup>+</sup> 188.1070, found 188.1073.

*Ethyl 2-Hydroxy-5-(5,6,7,8-tetrahydroisoquinolin-3-yl)pent-4-ynoate (18)*. By the general Sonogashira coupling procedure, 5,6,7,8-tetrahydroisoquinolin-3-yl triflate (3) (160 mg, 0.569 mmol), ethyl 2-hydroxypent-4-ynoate (17) (97 mg, 0.683 mmol), PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> (12 mg, 0.0171 mmol), and CuI (6.5 mg, 0.0341 mmol) in Et<sub>3</sub>N (1 mL) at 80 °C for 2 h provided 18 (145 mg, 94%) as an oil after flash chromatography (SiO<sub>2</sub>, hexanes/EtOAc = 1:1): IR (neat cm<sup>-1</sup>) 2934, 1734, 1195, 1098, 1033; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.13 (s, 1H), 6.98 (s, 1H), 4.64 (br, 1H), 4.41 (t, *J* = 5.6 Hz, 1H), 4.23–4.15 (m, 2H), 2.92 (qd, *J* = 16.8 and 4.8 Hz, 2H), 2.63–2.58 (m, 4H), 1.74–1.68 (m, 4H), 1.24 (t, *J* = 7.2 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 173.1, 150.3, 146.7, 139.5, 132.8, 127.3, 84.5, 82.7, 69.3, 61.8, 28.5, 26.3, 26.1, 22.5, 22.3, 14.3; HRMS (ES) *m/e* calcd for C<sub>16</sub>H<sub>20</sub>NO<sub>3</sub> (M + H)<sup>+</sup> 274.1438, found 274.1439.

*Ethyl 2-Hydroxy-3-(2-(5,6,7,8-tetrahydroisoquinolin-3-yl)-1H-indol-3-yl)propanoate (19)*. By the general Larock indole synthesis procedure, 2-bromoaniline (6) (102 mg, 0.593 mmol), alkyne 18 (135 mg, 0.494 mmol), Pd(OAc)<sub>2</sub> (5.6 mg, 0.0247 mmol), 1,1'-bis(diphenylphosphino)ferrocene (27.4 mg, 0.0494 mmol), and KHCO<sub>3</sub> (148 mg, 1.482 mmol) in DMF (2.5 mL) for 2 h provided 19 (120 mg, 67%) as an oil after flash chromatography (SiO<sub>2</sub>, hexanes/EtOAc = 1:1): IR (neat cm<sup>-1</sup>) 2929, 1729, 1603, 1205, 739; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.28 (s, 1H), 7.96 (s, 1H), 7.43–7.40 (m, 1H), 7.16 (s, 1H), 7.06–7.03 (m, 1H), 6.99–6.94 (m, 2H), 4.66 (t, *J* = 5.6 Hz, 1H), 4.36–4.28 (m, 1H), 4.21–4.13 (m, 1H), 3.45 (d, *J* = 5.6 Hz, 2H), 2.74–2.59 (m, 4H), 1.82–1.71 (m, 4H), 1.39 (t, *J* = 7.2 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 175.8, 148.2, 147.1, 146.6, 136.6, 134.6, 131.8, 128.9, 122.9, 121.7, 119.4, 118.5, 112.1, 110.1, 73.0, 61.3, 29.3, 29.1, 26.4, 22.8, 22.6, 14.6; HRMS (ES) *m/e* calcd for C<sub>22</sub>H<sub>25</sub>N<sub>2</sub>O<sub>3</sub> (M + H)<sup>+</sup> 365.1860, found 365.1861.

*7-(Ethoxycarbonyl)-2,3,4,7,8,13-hexahydro-1H-indolo[2',3':3,4]-pyridol[1,2-b]isoquinolin-6-ium Triflate (20)*. By the general triflate cyclization procedure, compound 19 (110 mg, 0.302 mmol), Et<sub>3</sub>N (46 mg, 0.453 mmol), and trifluoromethanesulfonic anhydride (62 μL, 0.362 mmol) in dry CHCl<sub>3</sub> (2.0 mL) at room temperature for 1 h provided 20 (119 mg, 80%) as a yellow solid: IR (neat cm<sup>-1</sup>) 3228, 1744, 1156, 1029, 768; <sup>1</sup>H NMR (400 MHz, DMSO) δ 12.32 (s, 1H), 8.82 (s, 1H), 8.13 (s, 1H), 7.81 (d, *J* = 8.0 Hz, 1H), 7.57 (d, *J* = 8.4 Hz, 1H), 7.41 (t, *J* = 7.2 Hz, 1H), 7.22 (t, *J* = 7.6 Hz, 1H), 6.14 (d, *J* = 6.0 Hz, 1H), 4.17–4.06 (m, 2H), 3.94 (d, *J* = 17.2 Hz, 1H), 3.74 (dd, *J* = 17.2 and 6.8 Hz, 1H), 3.19–3.06 (m, 2H), 2.98–2.83 (m, 2H), 1.97–1.83 (m, 4H), 1.10 (t, *J* = 6.8 Hz, 3H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 169.0, 160.3, 146.1, 140.1, 140.0, 134.7, 127.0, 125.7, 125.5, 121.8, 121.4, 121.3, 114.3, 113.6, 66.9, 63.7, 30.1, 26.5, 23.3, 21.8, 21.7, 14.7; HRMS (ES) *m/e* calcd for C<sub>22</sub>H<sub>23</sub>N<sub>2</sub>O<sub>2</sub> (M - OTf)<sup>+</sup> 347.1760, found 347.1763.

*7-(Ethoxycarbonyl)-2,3,4,13-tetrahydro-1H-indolo[2',3':3,4]-pyridol[1,2-b]isoquinolin-6-ium Triflate (21)*. By the general DDQ oxidation procedure, 20 (50 mg, 0.1 mmol) and 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (34 mg, 0.15 mmol) in acetic acid (1.0 mL) at 100 °C for 8 h provided 21 (40.2 mg, 82%) as a yellow solid: IR (neat cm<sup>-1</sup>) 2938, 1722, 1220, 1158, 756; <sup>1</sup>H NMR (400 MHz, DMSO) δ 13.42 (s, 1H), 9.88 (s, 1H), 9.28 (s, 1H), 8.59 (s, 1H), 8.46 (d, *J* = 8.0 Hz, 1H), 7.76–7.69 (m, 2H), 7.51 (td, *J* = 8.0 and 1.2 Hz, 1H), 4.65 (q, *J* = 7.2 Hz, 2H), 3.16 (br, 2H), 3.05 (br, 2H), 2.00–1.93 (m, 4H), 1.59 (t, *J* = 7.2 Hz, 3H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 162.6, 151.4, 141.6, 134.8, 134.6, 132.6, 131.9, 130.1, 124.2, 123.4,

123.3, 122.9, 122.1, 121.2, 119.6, 113.8, 63.9, 29.6, 27.3, 22.1, 22.0, 14.9; HRMS (ES)  $m/e$  calcd for  $C_{22}H_{21}N_2O_2$  ( $M - OTf$ )<sup>+</sup> 345.1603, found 345.1601.

**9-Methoxy-2,3,4,7,8,13-hexahydro-1H-indolo[2',3':3,4]pyrido[1,2-*b*]isoquinolin-6-ium Triflate (27).** A solution of 2-bromo-3-methoxyaniline (23) (53 mg, 0.26 mmol), alkyne 5 (41 mg, 0.2 mmol), Pd(OAc)<sub>2</sub> (4.5 mg, 0.02 mmol), 1,1'-bis(diphenylphosphino)ferrocene (22 mg, 0.04 mmol), and KHCO<sub>3</sub> (60 mg, 0.6 mmol) in DMF (1.0 mL) was degassed and stirred at 110 °C for 10 h. After being cooled to room temperature, water was added and the mixture was extracted with ethyl acetate. The organic extracts were combined, washed with water, filtered through a pad of silica gel, and dried to provide the crude intermediate 25. Et<sub>3</sub>N (31 mg, 0.3 mmol) and CHCl<sub>3</sub> (1.5 mL) were then added to this crude intermediate, and then trifluoromethanesulfonic anhydride (34 μL, 0.2 mmol) was slowly added to the mixture at 0 °C. After being stirred for 5 min, the resulting precipitate was filtered and washed with CHCl<sub>3</sub> to provide 27 (52 mg, 57%) as a yellow solid: IR (neat cm<sup>-1</sup>) 3246, 1513, 1247, 1032, 745; <sup>1</sup>H NMR (400 MHz, DMSO) δ 12.15 (s, 1H), 8.76 (s, 1H), 7.92 (s, 1H), 7.29 (t, *J* = 8.0 Hz, 1H), 7.11 (d, *J* = 8.4 Hz, 1H), 6.64 (d, *J* = 8.0 Hz, 1H), 4.79 (t, *J* = 6.8 Hz, 2H), 3.93 (s, 3H), 3.49 (t, *J* = 6.8 Hz, 2H), 3.01 (br, 2H), 2.84 (br, 2H), 1.85 (br, 4H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 158.1, 155.8, 145.1, 141.2, 140.1, 134.1, 128.0, 124.6, 120.7, 116.8, 116.6, 106.3, 101.3, 56.3, 55.8, 29.8, 26.4, 22.0, 21.9, 21.5; HRMS (ES)  $m/e$  calcd for  $C_{20}H_{21}N_2O$  ( $M - OTf$ )<sup>+</sup> 305.1654, found 305.1652.

**11-Methoxy-2,3,4,7,8,13-hexahydro-1H-indolo[2',3':3,4]pyrido[1,2-*b*]isoquinolin-6-ium Triflate (28).** A solution of 2-bromo-5-methoxyaniline (24) (51 mg, 0.25 mmol), 5 (51 mg, 0.25 mmol), Pd(OAc)<sub>2</sub> (5.6 mg, 0.025 mmol), 1,1'-bis(diphenylphosphino)ferrocene (28 mg, 0.05 mmol), and K<sub>2</sub>CO<sub>3</sub> (87 mg, 0.625 mmol) in DMF (1.5 mL) was degassed and stirred at 110 °C for 4 h. After being cooled to room temperature, water was added and the mixture was extracted with ethyl acetate. The organic extracts were combined, washed with water, filtered through a pad of silica gel, and dried to provide the crude intermediate 26. Et<sub>3</sub>N (38 mg, 0.375 mmol) and CHCl<sub>3</sub> (2 mL) were then added to this crude intermediate, and then trifluoromethanesulfonic anhydride (42 μL, 0.25 mmol) was slowly added to the mixture at 0 °C. After being stirred for 5 min, the resulting precipitate was filtered and washed with CHCl<sub>3</sub> to provide 28 (79 mg, 70%) as a yellow solid: IR (neat cm<sup>-1</sup>) 3293, 1554, 1249, 1031, 810; <sup>1</sup>H NMR (400 MHz, DMSO) δ 11.98 (s, 1H), 8.71 (s, 1H), 7.87 (s, 1H), 7.62 (d, *J* = 8.8 Hz, 1H), 6.95 (d, *J* = 2.0 Hz, 1H), 6.86 (dd, *J* = 8.8 and 2.0 Hz, 1H), 4.79 (t, *J* = 7.2 Hz, 2H), 3.88 (s, 3H), 3.34 (t, *J* = 7.2 Hz, 2H), 3.01 (br, 2H), 2.84 (br, 2H), 1.85 (br, 4H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 159.8, 157.9, 144.9, 141.2, 140.3, 133.5, 124.8, 122.2, 120.3, 120.2, 117.8, 112.9, 95.1, 56.3, 55.7, 29.8, 26.3, 22.0, 21.9, 19.9; HRMS (ES)  $m/e$  calcd for  $C_{20}H_{21}N_2O$  ( $M - OTf$ )<sup>+</sup> 305.1654, found 305.1653.

**9-Methoxy Sempervirine (29).** By the general DDQ oxidation procedure, 27 (46 mg, 0.1 mmol) and 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (30 mg, 0.13 mmol) in acetic acid (1.0 mL) at 100 °C for 30 min provided 29 (42 mg, 93%) as a yellow solid: IR (neat cm<sup>-1</sup>) 3171, 1352, 1240, 1026, 733; <sup>1</sup>H NMR (400 MHz, DMSO) δ 13.17 (s, 1H), 9.21 (s, 1H), 8.78 (d, *J* = 6.8 Hz, 1H), 8.52 (s, 1H), 8.41 (d, *J* = 6.8 Hz, 1H), 7.66 (t, *J* = 8.0 Hz, 1H), 7.37 (d, *J* = 8.0 Hz, 1H), 6.96 (d, *J* = 8.0 Hz, 1H), 4.12 (s, 3H), 3.17–3.12 (m, 2H), 3.02–2.98 (m, 2H), 1.97–1.89 (m, 4H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 156.8, 149.7, 143.0, 135.5, 133.5, 131.4, 131.0, 129.8, 127.2, 121.3, 120.0, 117.9, 111.7, 106.1, 102.8, 56.8, 29.8, 26.8, 22.2, 22.2; HRMS (ES)  $m/e$  calcd for  $C_{20}H_{19}N_2O$  ( $M - OTf$ )<sup>+</sup> 303.1497, found 303.1497.

**11-Methoxy Sempervirine (30).** By the general DDQ oxidation procedure, 28 (68 mg, 0.15 mmol) and 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (52 mg, 0.225 mmol) in acetic acid (1.0 mL) at 100 °C for 2 h provided 30 (64 mg, 94%) as a yellow solid: IR (neat cm<sup>-1</sup>) 1626, 1251, 1158, 1033, 799; <sup>1</sup>H NMR (400 MHz, DMSO) δ 13.06 (s, 1H), 9.15 (s, 1H), 8.81 (d, *J* = 5.2 Hz, 1H), 8.57 (d, *J* = 6.0 Hz, 1H), 8.52 (s, 1H), 8.24 (d, *J* = 8.8 Hz, 1H), 7.16 (s, 1H), 7.08 (d, *J* = 8.4 Hz, 1H), 3.98 (s, 3H), 3.15 (br, 2H), 3.00 (br, 2H), 1.93 (br, 4H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 161.8, 149.4, 143.4, 135.2, 133.0, 130.9,

130.1, 127.1, 123.7, 122.7, 119.8, 116.2, 115.5, 113.2, 95.4, 56.6, 29.7, 26.7, 22.2, 22.2; HRMS (ES)  $m/e$  calcd for  $C_{20}H_{19}N_2O$  ( $M - OTf$ )<sup>+</sup> 303.1497, found 303.1500.

**7,8-Dihydro-12-fluorosempervirine (35).** A solution of 2-bromo-6-fluoroaniline (31) (46 mg, 0.24 mmol), alkyne 5 (41 mg, 0.2 mmol), Pd(OAc)<sub>2</sub> (2.3 mg, 0.01 mmol), 1,1'-bis(diphenylphosphino)ferrocene (11 mg, 0.02 mmol), and KHCO<sub>3</sub> (60 mg, 0.6 mmol) in DMF (1.0 mL) was degassed and stirred at 110 °C for 2 h. After being cooled to room temperature, water was added and the mixture was extracted with ethyl acetate. The organic extracts were combined, washed with water, filtered through a pad of silica gel, and dried to provide the crude intermediate 33. Et<sub>3</sub>N (31 mg, 0.3 mmol) and CHCl<sub>3</sub> (1.5 mL) were then added to this crude intermediate, and then trifluoromethanesulfonic anhydride (34 μL, 0.2 mmol) was slowly added to the mixture at 0 °C. After being stirred for 5 min, the resulting precipitate was filtered and washed with CHCl<sub>3</sub> to provide 35 (65 mg, 74%) as a yellow solid: IR (neat cm<sup>-1</sup>) 3247, 1557, 1239, 1029, 787; <sup>1</sup>H NMR (400 MHz, DMSO) δ 12.58 (s, 1H), 8.84 (s, 1H), 8.15 (s, 1H), 7.58 (d, *J* = 8.0 Hz, 1H), 7.25–7.14 (m, 2H), 4.86 (t, *J* = 7.2 Hz, 2H), 3.40 (t, *J* = 7.2 Hz, 2H), 3.03 (br, 2H), 2.88 (br, 2H), 1.86 (br, 4H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 158.4, 151.4 (d, <sup>1</sup>*J*<sub>C-F</sub> = 243.5 Hz), 145.6, 139.8, 135.1, 129.4, 129.3, 127.9, 127.7, 127.1, 121.9, 121.9, 121.5, 117.5, 117.5, 117.5, 111.0, 110.9, 56.0, 29.9, 26.5, 21.9, 21.8, 19.8; HRMS (ES)  $m/e$  calcd for  $C_{19}H_{18}FN_2$  ( $M - OTf$ )<sup>+</sup> 293.1454, found 293.1450.

**7,8-Dihydro-10-fluorosempervirine (36).** A solution of alkyne 5 (70 mg, 0.348 mmol), Pd(OAc)<sub>2</sub> (2 mg, 0.0087 mmol), 1,1'-bis(diphenylphosphino)ferrocene (10 mg, 0.0174 mmol), and KHCO<sub>3</sub> (104 mg, 1.04 mmol) in DMF (1.6 mL) was degassed and stirred at 110 °C. A solution of 2-bromo-4-fluoroaniline (32) (199 mg, 1.04 mmol) in DMF (0.4 mL) was then added slowly to this solution, and the mixture was maintained at 110 °C for 4 h. After being cooled to room temperature, the resulting mixture was quenched with water and extracted with ethyl acetate. The organic extracts were combined, washed with water, filtered through a pad of silica gel, and dried to provide the crude intermediate. Et<sub>3</sub>N (53 mg, 0.522 mmol) and CHCl<sub>3</sub> (1 mL) were then added to this crude intermediate, and trifluoromethanesulfonic anhydride (60 μL, 0.348 mmol) was then added slowly to this mixture at 0 °C. After being stirred for 10 min, the resulting precipitate was filtered and washed with CHCl<sub>3</sub> to provide 36 (115 mg, 75%) as a yellow solid: IR (neat cm<sup>-1</sup>) 3285, 1561, 1282, 1034, 813; <sup>1</sup>H NMR (400 MHz, DMSO) δ 12.28 (s, 1H), 8.80 (s, 1H), 7.96 (s, 1H), 7.57–7.53 (m, 2H), 7.24 (td, *J* = 9.2 and 2.0 Hz, 1H), 4.84 (t, *J* = 7.2 Hz, 2H), 3.36 (t, *J* = 7.2 Hz, 2H), 3.03 (br, 2H), 2.87 (br, 2H), 1.86 (br, 4H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 159.5 (d, <sup>1</sup>*J*<sub>C-F</sub> = 233.4 Hz), 158.3, 145.5, 139.9, 136.4, 134.9, 127.6, 125.9, 125.8, 121.2, 116.7, 116.7, 115.4, 115.1, 114.8, 114.7, 105.8, 105.6, 56.0, 29.9, 26.5, 21.9, 21.8, 19.8; HRMS (ES)  $m/e$  calcd for  $C_{19}H_{18}FN_2$  ( $M - OTf$ )<sup>+</sup> 293.1454, found 293.1450.

**12-Fluorosempervirine (37).** By the general DDQ oxidation procedure, 35 (74 mg, 0.167 mmol) and 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (76 mg, 0.335 mmol) in acetic acid (1.8 mL) at 100 °C for 4 h provided 37 (66 mg, 90%) as a yellow solid: IR (neat cm<sup>-1</sup>) 2948, 1406, 1222, 1026, 776; <sup>1</sup>H NMR (400 MHz, DMSO) δ 13.40 (s, 1H), 9.17 (s, 1H), 8.79 (d, *J* = 7.2 Hz, 1H), 8.65 (s, 1H), 8.61 (d, *J* = 7.2 Hz, 1H), 8.15 (d, *J* = 7.6 Hz, 1H), 7.58 (dd, *J* = 11.2 and 8.0 Hz, 1H), 7.44–7.39 (m, 1H), 3.13 (br, 2H), 3.00 (br, 2H), 1.94 (br, 4H); <sup>13</sup>C NMR (100 MHz, DMSO) δ 151.0 (d, <sup>1</sup>*J*<sub>C-F</sub> = 244.5 Hz), 150.7, 136.0, 134.4, 131.7, 131.0, 129.6, 129.5, 127.5, 125.1, 125.1, 123.0, 122.9, 121.9, 121.8, 120.5, 118.8, 116.9, 114.3, 114.2, 29.8, 26.8, 22.13, 22.09; HRMS (ES)  $m/e$  calcd for  $C_{19}H_{16}FN_2$  ( $M - OTf$ )<sup>+</sup> 291.1298, found 291.1299.

**10-Fluorosempervirine (38).** By the general DDQ oxidation procedure, 36 (66 mg, 0.15 mmol) and 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (340 mg, 1.5 mmol) in acetic acid (1.0 mL) at 100 °C for 7 days provided 38 (34 mg, 52%) as a yellow solid: IR (neat cm<sup>-1</sup>) 2943, 1651, 1248, 1153, 820; <sup>1</sup>H NMR (400 MHz, DMSO) δ 13.66 (br, 1H), 9.25 (s, 1H), 8.82 (d, *J* = 6.8 Hz, 1H), 8.76 (s, 1H), 8.62 (d, *J* = 6.8 Hz, 1H), 8.24 (dd, *J* = 8.8 and 2.0 Hz, 1H), 7.85 (dd, *J* = 8.8 and 4.0 Hz, 1H), 7.60 (td, *J* = 9.2 and 2.0 Hz, 1H), 3.17 (br, 2H), 3.03 (br,

2H), 1.94 (br, 4H);  $^{13}\text{C}$  NMR (100 MHz, DMSO)  $\delta$  159.6 (d,  $^1J_{\text{C-F}}$  = 234.9 Hz), 150.5, 138.4, 136.0, 134.2, 131.9, 131.8, 126.7, 122.1, 122.0, 121.6, 121.5, 120.8, 118.3, 118.1, 117.0, 115.3, 115.2, 107.8, 107.6, 29.8, 26.8, 22.2, 22.1; HRMS (ES)  $m/e$  calcd for  $\text{C}_{19}\text{H}_{16}\text{FN}_2$  ( $\text{M} - \text{OTf}$ ) $^+$  291.1298, found 291.1295.

**13H-Indolo[2',3':3,4]pyrido[1,2-b]isoquinolin-6-ium Triflate (43).** By the general DDQ oxidation procedure, **42**<sup>2</sup> (42 mg, 0.1 mmol) and 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (34 mg, 0.15 mmol) in acetic acid (1.0 mL) at 100 °C for 30 min provided **43** (33 mg, 79%) as a yellow solid: IR (neat  $\text{cm}^{-1}$ ) 3215, 1221, 1027, 885, 733;  $^1\text{H}$  NMR (400 MHz, DMSO)  $\delta$  13.40 (s, 1H), 10.48 (s, 1H), 9.44 (s, 1H), 9.04 (d,  $J$  = 6.0 Hz, 1H), 8.70 (d,  $J$  = 6.4 Hz, 1H), 8.44 (d,  $J$  = 8.0 Hz, 1H), 8.34 (d,  $J$  = 10.0 Hz, 1H), 8.32 (d,  $J$  = 8.4 Hz, 1H), 8.15 (t,  $J$  = 7.2 Hz, 1H), 8.00 (t,  $J$  = 7.2 Hz, 1H), 7.78 (d,  $J$  = 8.4 Hz, 1H), 7.62 (t,  $J$  = 7.2 Hz, 1H), 7.42 (t,  $J$  = 7.6 Hz, 1H);  $^{13}\text{C}$  NMR (100 MHz, DMSO)  $\delta$  141.3, 140.7, 135.5, 135.4, 131.2, 130.9, 130.0, 129.3, 128.8, 127.9, 126.8, 125.8, 122.4, 122.2, 122.1, 119.4, 118.2, 117.7, 113.5; HRMS (ES)  $m/e$  calcd for  $\text{C}_{19}\text{H}_{13}\text{N}_2$  ( $\text{M} - \text{OTf}$ ) $^+$  269.1079, found 269.1077.

## ■ ASSOCIATED CONTENT

### ■ Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.6b00022.

$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra for new compounds (PDF)

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### Notes

The authors declare no competing financial interest.

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## ■ DEDICATION

This paper is dedicated to the memory of John Michael "Mike" Kane, an inspirational organic and medicinal chemist, mentor, and friend to T.D.B.

## ■ REFERENCES

- (1) Newman, D. J.; Cragg, G. M. *J. Nat. Prod.* **2012**, *75*, 311.
- (2) Pan, X.; Bannister, T. D. *Org. Lett.* **2014**, *16*, 6124.
- (3) (a) Sonogashira, K.; Tohda, Y.; Hagihara, N. *Tetrahedron Lett.* **1975**, *16*, 4467. (b) Chinchilla, R.; Nájera, C. *Chem. Soc. Rev.* **2011**, *40*, 5084.
- (4) (a) Larock, R. C.; Yum, E. K. *J. Am. Chem. Soc.* **1991**, *113*, 6689. (b) Larock, R. C.; Yum, E. K.; Refvik, M. D. *J. Org. Chem.* **1998**, *63*, 7652.
- (5) Woodward, R. B.; Witkop, B. *J. Am. Chem. Soc.* **1949**, *71*, 379.
- (6) Woodward, R. B.; McLamore, W. M. *J. Am. Chem. Soc.* **1949**, *71*, 379.
- (7) Swan, G. A. *J. Chem. Soc.* **1958**, 2038.
- (8) Westphal, O.; Jann, K.; Heffé, W. *Arch. Pharm.* **1961**, *294*, 37.
- (9) Potts, K. T.; Mattingly, G. S. *J. Org. Chem.* **1968**, *33*, 3985.
- (10) (a) Gribble, G. W.; Johnson, D. A. *Tetrahedron Lett.* **1987**, *28*, 5259. (b) Gribble, G. W.; Barden, T. C.; Johnson, D. A. *Tetrahedron* **1988**, *44*, 3195.
- (11) (a) Honda, R.; Tanaka, Yasuda, H. H. *FEBS Lett.* **1997**, *420*, 25. (b) Sasiela, C. A.; Stewart, D. H.; Kitagaki, J.; Safiran, Y. J.; Yang, Y.; Weissman, Y. A.; Oberoi, P.; Davydov, I. V.; Goncharova, E.; Beutler, J. A.; McMahon, J. B.; O'Keefe, B. R. *J. Biomol. Screening* **2008**, *13*, 229. (c) Matlashewski, G.; Lamb, P.; Pim, D.; Peacock, J.; Crawford, L.; Benchimol, S. *EMBO J.* **1984**, *3*, 3257. (d) Peters, J. M.; Franke, W.

W.; Kleinschmidt, J. A. *J. Biol. Chem.* **1994**, *269*, 7709. (e) Dickens, P.; Fitzgerald, R.; Fischer, P. M. *Semin. Cancer Biol.* **2010**, *20*, 10. (f) Zhang, Z.; Wang, P.; Yuan, W.; Li, S. *Planta Med.* **2008**, *74*, 1818. (g) Beljanski, M.; Beljanski, M. S. *Oncology* **1986**, *43*, 198.

(12) (a) Lipińska, T. M. *Tetrahedron* **2005**, *61*, 8148. (b) Lipińska, T. M. *Tetrahedron Lett.* **2002**, *43*, 9565. (c) Lipińska, T. M. *Tetrahedron Lett.* **2004**, *45*, 8831. (d) Lipińska, T. M.; Czarnocki, S. J. *Org. Lett.* **2006**, *8*, 367.

(13) Chinta Rao, T. S.; Saha, S.; Raolji, G. B.; Patro, B.; Risbood, P.; Difilippantonio, M. J.; Tomaszewski, J. E.; Malhotra, S. V. *Tetrahedron Lett.* **2013**, *54*, 487.

(14) Huebner, C. F.; St. Andre, A. F.; Schlittler, E.; Uffer, A. *J. Am. Chem. Soc.* **1955**, *77*, 5725.

(15) (a) Zhang, Y.; Selley, D. E.; Dewey, W. Patent Appl. WO 2010083384 A3, 2010. (b) Koltunov, K. Y.; Prakash, G. K. S.; Rasul, G.; Olah, G. A. *J. Org. Chem.* **2002**, *67*, 8943.

(16) Dyker, G.; Hildebrandt, D. *J. Org. Chem.* **2005**, *70*, 6093.

(17) Products of the Larock reactions reported in this study (compounds **7**, **11**, **19**, **25**, **26**, **33**, **34**, and **41**) were obtained with  $\geq 95\%$  regioselectivity, as no isomer was detected by  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopy. In a few instances (**11**, **25**, **26**, **41**), there was analytical HPLC/LCMS evidence for the formation of trace amounts of a regioisomer (estimated yields ranging from 1 to 3%), but in no instances were these minor isomers isolated for full characterization.

(18) Doherty, J. R.; Yang, C.; Scott, K. E. N.; Cameron, M. D.; Fallahi, M.; Li, W.; Hall, M. A.; Amelio, A. L.; Mishra, J. K.; Li, F.; Tortosa, M.; Genau, H. M.; Rounbehler, R. J.; Lu, Y.; Dang, C. V.; Kumar, K. G.; Butler, A. A.; Bannister, T. D.; Hooper, A. T.; Unsal-Kacmaz, K.; Roush, W. R.; Cleveland, J. L. *Cancer Res.* **2014**, *74*, 908.

(19) In comparable assays, the marketed topoisomerase I inhibitor topotecan has 2–5-fold higher potency, suggesting that additional rounds of optimization of compounds **29** and **38** may lead to compounds of significant activity. For the topotecan data, see: Mitsui, I.; Kumazawa, E.; Hirota, Y.; Aonuma, M.; Sugimori, M.; Ohsuki, S.; Uoto, K.; Ejima, A.; Terasawa, H.; Sato, K. *Jpn. J. Cancer Res.* **1995**, *86*, 776.